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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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<p>(54) Title: 1,3-OXATHIOLANE NUCLEOSIDE ANALOGUES</p>		
<p>(57) Abstract</p> <p>(-)-4-Amino-1-(2-hydroxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-one, its pharmaceutically acceptable derivatives, pharmaceutical formulations thereof, methods for its preparation and its use as an antiviral agent are described.</p>		

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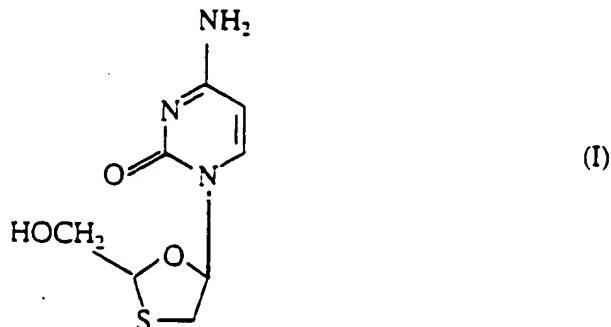
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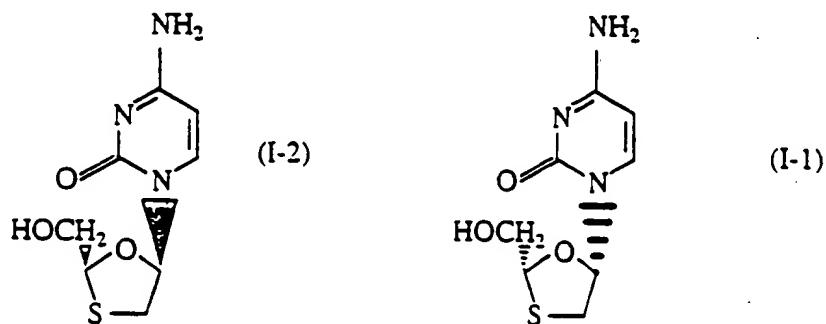
1,3-Oxathiolane Nucleoside Analogues

The present invention relates to nucleoside analogues and their use in medicine. More specifically the invention is concerned with 1,3-oxathiolane nucleoside analogues, pharmaceutical formulations thereof and the use thereof in the treatment of viral infections.

The compound of formula (I)



also known as BCH-189 or NGPB-21 has been described as having antiviral activity in particular against the human immunodeficiency viruses (HIV's), the causative agents of AIDS (5th Anti-Aids Conference, Montreal, Canada 5th-9th June 1989: Abstracts T.C.O.1 and M.C.P. 63; European Patent Application Publication No. 0382562). The compound of formula (I) is a racemic mixture of the two enantiomers of formulae (I-1) and (I-2):-



and was described and tested in the form of its racemate. The only compound currently approved for the treatment of conditions caused by HIV is 3'-azido-3'-deoxythymidine (AZT, zidovudine, BW 509U). However, this compound has a significant side-effect liability and thus either cannot be employed or, once employed, may have to be withdrawn in a significant number of patients. There is in

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consequence a continuing need to provide compounds which are effective against HIV but with a concomitant significantly better therapeutic index.

We have now found that, surprisingly, enantiomers of the compound of formula (I) are equipotent against HIV but that one of the enantiomers (the (-)-enantiomer) has considerably lower cytotoxicity than the other enantiomer. There is thus provided in a first aspect of the invention the (-) (or laevorotatory) enantiomer of the compound of formula (I) and pharmaceutically acceptable derivatives thereof.

The (-) enantiomer has the chemical name (-)-cis-4-Amino-1-(2-hydroxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-one (hereinafter compound (A)). It has the absolute stereochemistry of the compound of formula (I-1) which has the name (2R,cis))-4-amino-1-(2-hydroxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-one.

Preferably Compound A is provided substantially free of the corresponding (+)-enantiomer, that is to say no more than about 5% w/w of the (+)- enantiomer, preferably no more than about 2%, in particular less than about 1% w/w is present.

By "a pharmaceutically acceptable derivative" is meant any pharmaceutically acceptable salt, ester, or salt of such ester, of compound (A) or any other compound which, upon administration to the recipient, is capable of providing (directly or indirectly) compound (A) or an antivirally active metabolite or residue thereof.

It will be appreciated by those skilled in the art that compound (A) may be modified to provide pharmaceutically acceptable derivatives thereof, at functional groups in both the base moiety and at the hydroxymethyl group of the oxathiolane ring. Modification at all such functional groups are included within the scope of the invention. However of particular interest are pharmaceutically acceptable derivatives obtained by modification of the 2-hydroxymethyl group of the oxathiolane ring.

Preferred esters of compound (A) include the compounds in which the hydrogen of the 2-hydroxymethyl group is replaced by an acyl

function $R-\overset{O}{\underset{H}{\overset{|}{C}}}-$ in which the non-carbonyl moiety R of the ester is selected from hydrogen, straight or branched chain alkyl (e.g.

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methyli, ethyli, n-propyl, t-butyli, n-butyli), alkoxyalkyli (e.g. methoxymethyl), aralkyl (e.g. benzyl), aryloxyalkyl (e.g. phenoxyethyl), aryl (e.g. phenyl optionally substituted by halogen, C₁-4 alkyl or C₁-4 alkoxy); sulphonate esters such as alkyl- or aralkylsulphonyli (e.g. methanesulphonyli); amino acid esters (e.g. L-valyl or L-isoleucyl) and mono-, di- or tri-phosphate esters.

With regard to the above described esters, unless otherwise specified, any alkyl moiety present advantageously contains 1 to 16 carbon atoms, particularly 1 to 4 carbon atoms. Any aryl moiety present in such esters advantageously comprises a phenyl group.

In particular the esters may be a C₁-16alkyl ester, an unsubstituted benzyl ester or a benzyl ester substituted by at least one halogen (bromine, chlorine, fluorine or iodine), C₁-6alkyl, C₁-6alkoxy, nitro or trifluoromethyl groups.

Pharmaceutically acceptable salts of the compound (A) include those derived from pharmaceutically acceptable inorganic and organic acids and bases. Examples of suitable acids include hydrochloric, hydrobromic, sulphuric, nitric, perchloric, fumaric, maleic, phosphoric, glycollic, lactic, salicylic, succinic, toluene-p-sulphonic, tartaric, acetic, citric, methanesulphonic, formic, benzoic, malonic, naphthalene-2-sulphonic and benzenesulphonic acids. Other acids such as oxalic, while not in themselves pharmaceutically acceptable, may be useful as intermediates in obtaining the compounds of the invention and their pharmaceutically acceptable acid addition salts.

Salts derived from appropriate bases include alkali metal (e.g. sodium), alkaline earth metal (e.g. magnesium), ammonium and NR₄⁺ (where R is C₁-4alkyl) salts.

References hereinafter to a compound according to the invention include both the compound (A) and its pharmaceutically acceptable derivatives.

The compounds of the invention either themselves possess antiviral activity and/or are metabolizable to such compounds. In particular these compounds are effective in inhibiting the replication of retroviruses, including human retroviruses such as human immunodeficiency viruses (HIV's), the causative agents of AIDS.

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There is thus provided as a further aspect of the invention compound (A) or a pharmaceutically acceptable derivative thereof for use as an active therapeutic agent in particular as an antiviral agent, for example in the treatment of retroviral infections.

In a further or alternative aspect there is provided a method for the treatment of a viral infection, in particular an infection caused by a retrovirus such as HIV, in a mammal including man comprising administration of an effective amount of compound (A) or a pharmaceutically acceptable derivative thereof.

There is also provided in a further or alternative aspect use of compound (A) or a pharmaceutically acceptable derivative thereof for the manufacture of a medicament for the treatment of a viral infection.

The compounds of the invention are also useful in the treatment of AIDS related conditions such as AIDS-related complex (ARC), progressive generalised lymphadenopathy (PGL), AIDS-related neurological conditions (such as dementia or tropical paraparesis), anti-HIV antibody positive and HIV- positive conditions, Kaposi's sarcoma, thrombocytopenia purpura and associated opportunistic infections for example *Pneumocystis carinii*.

The compounds of the invention are also useful in the prevention of progression to clinical illness of individuals who are anti-HIV antibody or HIV-antigen positive and in prophylaxis following exposure to HIV.

The compound (A) or pharmaceutically acceptable derivatives thereof may also be used for the prevention of viral contamination of physiological fluids such as blood or semen in vitro.

It will be appreciated by those skilled in the art that reference herein to treatment extends to prophylaxis as well as the treatment of established infections or symptoms.

It will be further appreciated that the amount of a compound of the invention required for use in treatment will vary not only with the particular compound selected but also with the route of administration, the nature of the condition being treated and the age and condition of the patient and will be ultimately at the discretion of the attendant physician or veterinarian. In general however a suitable dose will be in the range of from about 0.1 to

about 750mg/kg of bodyweight per day preferably in the range of 0.5 to 60 mg/kg/day, most preferably in the range of 1 to 20mg/kg/day.

The desired dose may conveniently be presented in a single dose or as divided doses administered at appropriate intervals, for example as two, three, four or more sub-doses per day.

The compound is conveniently administered in unit dosage form, for example containing 10 to 1500mg, conveniently 20 to 1000mg, most conveniently 50 to 700mg of active ingredient per unit dosage form.

Ideally the active ingredient should be administered to achieve peak plasma concentrations of the active compound of from about 1 to about 75 μ M, preferably about 2 to 50 μ M, most preferably about 3 to about 30 μ M. This may be achieved, for example, by the intravenous injection of a 0.1 to 5% solution of the active ingredient, optionally in saline, or orally administered as a bolus containing about 1 to about 100mg of the active ingredient. Desirable blood levels may be maintained by a continuous infusion to provide about 0.01 to about 5.0 mg/kg/hour or by intermittent infusions containing about 0.4 to about 15 mg/kg of the active ingredient.

While it is possible that, for use in therapy, a compound of the invention may be administered as the raw chemical it is preferable to present the active ingredient as a pharmaceutical formulation.

The invention thus further provides a pharmaceutical formulation comprising compound (A) or a pharmaceutically acceptable derivative thereof together with one or more pharmaceutically acceptable carriers therefor and, optionally, other therapeutic and/or prophylactic ingredients. The carrier(s) must be 'acceptable' in the sense of being compatible with the other ingredients of the formulation and not deleterious to the recipient thereof.

Pharmaceutical formulations include those suitable for oral, rectal, nasal, topical (including buccal and sub-lingual), vaginal or parenteral (including intramuscular, sub-cutaneous and intravenous) administration or in a form suitable for administration by inhalation or insufflation. The formulations may, where appropriate, be conveniently presented in discrete dosage units and may be prepared by any of the methods well known in the art of

pharmacy. All methods include the step of bringing into association the active compound with liquid carriers or finely divided solid carriers or both and then, if necessary, shaping the product into the desired formulation.

Pharmaceutical formulations suitable for oral administration may conveniently be presented as discrete units such as capsules, cachets or tablets each containing a predetermined amount of the active ingredient; as a powder or granules; as a solution, a suspension or as an emulsion. The active ingredient may also be presented as a bolus, electuary or paste. Tablets and capsules for oral administration may contain conventional excipients such as binding agents, fillers, lubricants, disintegrants, or wetting agents. The tablets may be coated according to methods well known in the art. Oral liquid preparations may be in the form of, for example, aqueous or oily suspensions, solutions, emulsions, syrups or elixirs, or may be presented as a dry product for constitution with water or other suitable vehicle before use. Such liquid preparations may contain conventional additives such as suspending agents, emulsifying agents, non-aqueous vehicles (which may include edible oils), or preservatives.

The compounds according to the invention may also be formulated for parenteral administration (e.g. by injection, for example bolus injection or continuous infusion) and may be presented in unit dose form in ampoules, pre-filled syringes, small volume infusion or in multi-dose containers with an added preservative. The compositions may take such forms as suspensions, solutions, or emulsions in oily or aqueous vehicles, and may contain formulatory agents such as suspending, stabilising and/or dispersing agents. Alternatively, the active ingredient may be in powder form, obtained by aseptic isolation of sterile solid or by lyophilisation from solution, for constitution with a suitable vehicle, e.g. sterile, pyrogen-free water, before use.

For topical administration to the epidermis the compounds according to the invention may be formulated as ointments, creams or lotions, or as a transdermal patch. Ointments and creams may, for example, be formulated with an aqueous or oily base with the addition of suitable thickening and/or gelling agents. Lotions may

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be formulated with an aqueous or oily base and will in general also contain one or more emulsifying agents, stabilising agents, dispersing agents, suspending agents, thickening agents, or colouring agents.

Formulations suitable for topical administration in the mouth include lozenges comprising active ingredient in a flavoured base, usually sucrose and acacia or tragacanth; pastilles comprising the active ingredient in an inert base such as gelatin and glycerin or sucrose and acacia; and mouthwashes comprising the active ingredient in a suitable liquid carrier.

Pharmaceutical formulations suitable for rectal administration wherein the carrier is a solid are most preferably presented as unit dose suppositories. Suitable carriers include cocoa butter and other materials commonly used in the art, and the suppositories may be conveniently formed by admixture of the active compound with the softened or melted carrier(s) followed by chilling and shaping in moulds.

Formulations suitable for vaginal administration may be presented as pessaries, tampons, creams, gels, pastes, foams or sprays containing in addition to the active ingredient such carriers as are known in the art to be appropriate.

For intra-nasal administration the compounds of the invention may be used as a liquid spray or dispersible powder or in the form of drops.

Drops may be formulated with an aqueous or non-aqueous base also comprising one or more dispersing agents, solubilising agents or suspending agents. Liquid sprays are conveniently delivered from pressurised packs.

For administration by inhalation the compounds according to the invention are conveniently delivered from an insufflator, nebuliser or a pressurised pack or other convenient means of delivering an aerosol spray. Pressurised packs may comprise a suitable propellant such as dichlorodifluoromethane, trichlorofluoromethane, dichlorotetrafluoroethane, carbon dioxide or other suitable gas. In the case of a pressurised aerosol the dosage unit may be determined by providing a valve to deliver a metered amount.

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Alternatively, for administration by inhalation or insufflation, the compounds according to the invention may take the form of a dry powder composition, for example a powder mix of the compound and a suitable powder base such as lactose or starch. The powder composition may be presented in unit dosage form in, for example, capsules or cartridges or e.g. gelatin or blister packs from which the powder may be administered with the aid of an inhalator or insufflator.

When desired the above described formulations adapted to give sustained release of the active ingredient may be employed.

The pharmaceutical compositions according to the invention may also contain other active ingredients such as antimicrobial agents, or preservatives.

The compounds of the invention may also be used in combination with other therapeutic agents for example other antiinfective agents. In particular the compounds of the invention may be employed together with known antiviral agents.

The invention thus provides, in a further aspect, a combination comprising the compound (A) or a physiologically acceptable derivative thereof together with another therapeutically active agent, in particular an antiviral agent.

The combinations referred to above may conveniently be presented for use in the form of a pharmaceutical formulation and thus pharmaceutical formulations comprising a combination as defined above together with a pharmaceutically acceptable carrier therefor comprise a further aspect of the invention.

Suitable therapeutic agents for use in such combinations include acyclic nucleosides such as acyclovir or ganciclovir, interferons such as α , β or γ -interferon, renal excretion inhibitors such as probenecid, nucleoside transport inhibitors such as dipyridamole, 2',3"-dideoxynucleosides such as AZT, 2',3"-dideoxycytidine, 2',3"-dideoxyadenosine, 2',3"-dideoxyinosine, 2',3"-dideoxythymidine, 2',3"-dideoxy-2',3"-didehydrothymidine and 2',3"-dideoxy-2',3"-didehydrocytidine, immunomodulators such as interleukin II (IL2) and granulocyte macrophage colony stimulating factor (GM-CSF), erythropoietin, ampligen, thymomulin, thym pentin, foscarnet, ribavirin and inhibitors of HIV binding to CD4 receptors

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e.g. soluble CD4, CD4 fragments, CD4 hybrid molecules, glycosylation inhibitors such as 2-deoxy-D-glucose, castanospermine and 1-deoxynojirimycin.

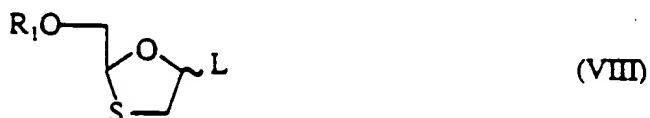
The individual components of such combinations may be administered either sequentially or simultaneously in separate or combined pharmaceutical formulations.

When the compound (A) or a pharmaceutically acceptable derivative thereof is used in combination with a second therapeutic agent active against the same virus the dose of each compound may be either the same as or differ from that when the compound is used alone. Appropriate doses will be readily appreciated by those skilled in the art.

The compound (A) and its pharmaceutically acceptable derivatives may be prepared by any method known in the art for the preparation of compounds of analogous structure, for example as described in European Patent Application Publication No. 0382562.

It will be appreciated by those skilled in the art that for certain of the methods described herein below the desired stereochemistry of the compound (A) may be obtained either by commencing with an optically pure starting material or by resolving the racemic mixture at any convenient stage in the synthesis. In the case of all the processes the optically pure desired product may be obtained by resolution of the end product of each reaction.

In one such process (A) a 1,3-oxathiolane of formula (VIII)

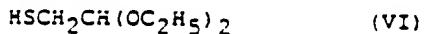


wherein the anomeric group L is a displaceable group, is reacted with an appropriate base. Suitable groups L include -OR where R is an alkyl group, e.g. a C₁-6alkyl group such as methyl or R is an acyl group, e.g. a C₁-6alkyl group such as acetyl or halogen, for example iodine, bromine or chlorine.

The compound of formula VIII is conveniently reacted with cytosine or an appropriate pyrimidine base precursor thereof (previously silylated with a silylating agent such as hexamethyldisilazane) in a compatible solvent such as methylene

chloride using a Lewis acid such as titanium tetrachloride, tin (IV) compound such as SnCl_4 , or trimethylsilyltriflate.

The 1,3-oxathiolanes of formula (VIII) may be prepared for example by reaction of an aldehyde of formula (VII) with a mercaptoacetal of formula (VI) in a compatible organic solvent, such as toluene in the presence of an acid catalyst for example a Lewis acid such as zinc chloride.



The mercaptoacetals of formula (VI) may be prepared by methods known in the art, for example G. Hesse and I. Jorder, Chem. Ber. 85, 924-932 (1952).

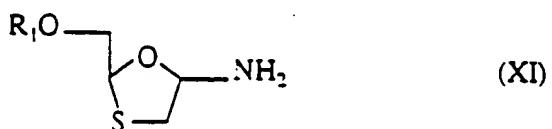
The aldehydes of formula (VII) may be prepared by methods known in the art for example E. G. Halloquist and H. Hibbert, Can. J. Research, 8, 129-136 (1933). Conveniently the crude aldehyde (VII) may be purified by conversion to the crystalline bisulphite addition adduct and subsequent reconversion to the free aldehyde.

In a second process (B) the compound (A) is obtained by base interconversion of a compound of formula (IX)



where B is a base convertible to cytosine. Such interconversion may be effected either by simple chemical transformation (e.g. the conversion of uracil base to cytosine) or by an enzymatic conversion using a deoxyribosyl transferase. Such methods and conditions for base interconversion are well known in the art of nucleoside chemistry.

In a third process (C) a compound of formula (XI)



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may be converted to the compound (A) by conversion of the anomeric NH₂ group to the cytosine base by methods well known in the nucleoside chemistry art.

Many of the reactions described hereinabove have been extensively reported in the context of nucleoside synthesis, for example in Nucleoside Analogs - Chemistry, Biology and Medical Applications, R. T. Walker *et. al.*, eds, Plenum Press, New York (1979) at pages 165-192; T. Ueda in Chemistry of Nucleosides and Nucleotides, Vol I, L B Townsend ed., Plenum Press, New York (1988) at pages 165-192 the disclosures of which are incorporated by reference herein.

It will be appreciated that the above reactions may require the use of, or conveniently may be applied to, starting materials having protected functional groups, and deprotection might thus be required as an intermediate or final step to yield the desired compound. Protection and deprotection of functional groups may be effected using conventional means. Thus, for example, amino groups may be protected by a group selected from aralkyl (e.g. benzyl), acyl, aryl (e.g. 2,4-dinitrophenyl) or silyl; subsequent removal of the protecting group being effected when desired by hydrolysis or hydrogenolysis as appropriate using standard conditions. Hydroxyl groups may be protected using any conventional hydroxyl protecting group, for example, as described in 'Protective Groups in Organic Chemistry', Ed. J. F. W. McOmie (Plenum Press, 1973) or 'Protective Groups in Organic Synthesis' by Theodora W. Greene (John Wiley and Sons, 1981). Examples of suitable hydroxyl protecting groups include groups selected from alkyl (e.g. methyl, *t*-butyl or methoxymethyl), aralkyl (e.g. benzyl, diphenylmethyl or triphenylmethyl), heterocyclic groups such as tetrahydropyranyl, acyl (e.g. acetyl or benzoyl) and silyl groups such as trialkylsilyl (e.g. *t*-butyldimethylsilyl). The hydroxyl protecting groups may be removed by conventional techniques. Thus, for example, alkyl, silyl, acyl and heterocyclic groups may be removed by solvolysis, e.g. by hydrolysis under acidic or basic conditions. Aralkyl groups such as triphenylmethyl may similarly be removed by solvolysis, e.g. by hydrolysis under acidic conditions. Aralkyl groups such as benzyl may be cleaved for example by treatment with BF₃/etherate and

acetic anhydride followed by removal of acetate groups so formed at an appropriate stage in the synthesis. Silyl groups may also conveniently be removed using a source of fluoride ions such as tetra-n-butylammonium fluoride.

In the above processes compound (A) is generally obtained as a mixture of the cis and trans isomers of which the cis isomer is the compound of interest.

These isomers may be separated by physical means e.g. chromatography on silica gel or by fractional crystallisation, either directly or on a suitable derivatives thereof, eg acetates (prepared for example with acetic anhydride) followed, after separation, by conversion back to the parent product (eg by deacetylation with methanolic ammonia).

Pharmaceutically acceptable salts of the compounds of the invention may be prepared as described in US Patent No. 4,383,114, the disclosure of which is incorporated by reference herein. Thus, for example, when it is desired to prepare an acid addition salt of compound (A) the product of any of the above procedures may be converted into a salt by treatment of the resulting free base with a suitable acid using conventional methods. Pharmaceutically acceptable acid addition salts may be prepared by reacting the free base with an appropriate acid optionally in the presence of a suitable solvent such as an ester (e.g. ethyl acetate) or an alcohol (e.g. methanol, ethanol or isopropanol). Inorganic basic salts may be prepared by reacting the parent compound with a suitable base such as an alkoxide (e.g. sodium methoxide) optionally in the presence of a solvent such as an alcohol (e.g. methanol). Pharmaceutically acceptable salts may also be prepared from other salts, including other pharmaceutically acceptable salts, of the compound (A) using conventional methods.

Compound (A) may be converted into a pharmaceutically acceptable phosphate or other ester by reaction with a phosphorylating agent, such as POCl_3 , or a suitable esterifying agent, such as an acid halide or anhydride, as appropriate. An ester or salt of compound (A) may be converted to the parent compound for example by hydrolysis.

Resolution of the final product, or an intermediate or starting material therefor may be effected by any suitable method known in the art : see for example 'Stereochemistry of Carbon Compounds' by E. L. Eliel (McGraw Hill, 1962) and 'Tables of Resolving Agents' by S. H. Wilen.

Thus for example the compound (A) may be obtained by chiral HPLC using a suitable stationary phase for example acetylated 3-cyclodextrin or cellulose triacetate and a suitable solvent for example an alcohol such as ethanol or an aqueous solution of for example triethyl ammonium acetate. Alternatively the compounds may be resolved by enzyme mediated enantioselective catabolism with a suitable enzyme such as cytidine deaminase or selective enzymatic degradation of a suitable derivative using a 5'-nucleotidase. When resolution is effected enzymatically the enzyme may be employed either in solution or, more conveniently, in immobilised form. Enzymes may be immobilised by any method known in the art, for example by adsorption onto a resin such as Supergit C.

The invention will be further described by the following examples which are not intended to limit the invention in any way. All temperatures are in degrees Celcius.

INTERMEDIATE 1

5-Methoxy-1,3-oxathiolane-2-methanol, benzoate.

A solution of zinc chloride (1.6g) in hot methanol (15ml) was added to a stirred solution of mercaptoacetaldehyde, dimethyl acetal (34.2g) and benzyloxy acetaldehyde (48.3g) in toluene (1300ml) which was then heated to reflux under nitrogen for 50 min. The cooled mixture was concentrated, diluted with some toluene, then filtered through Kieselguhr. The combined filtrates and toluene were washed with aqueous saturated sodium bicarbonate solution (x2) and brine, dried ($MgSO_4$) then evaporated to an oil which was subjected to column chromatography on silica (2kg, Merck 9385) eluted with chloroform to give the title product as an oil (45.1g) a mixture of anomers (ca 1:1); 1H NMR ($DMSO-d_6$) 3.1-3.3(4H), 3.42(6H), 4.4-4.6 (4H), 5.41(1H), 5.46 (1H), 5.54 (1H), 5.63 (1H), 7.46 (4H), 7.58 (2H), 8.07 (4H); γ_{max} ($CHBr_3$) 1717.6cm^{-1} .

INTERMEDIATE 2

(±)-cis-1-(2-Benzovloxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-4-dione

A mixture of finely ground uracil (9.62g) hexamethyl disilazane (50 ml) and ammonium sulphate (30 mg) was heated at reflux under nitrogen until a clear solution was obtained. This was cooled and then evaporated to a colourless oil, which was dissolved, under nitrogen atmosphere, in acetonitrile (100ml). The solution was added to a stirred ice cooled solution of 5-methoxy-1,3-oxathiolane-2-methanol, benzoate (Intermediate 1) (19.43g), in acetonitrile (600ml) and trimethyl silyl trifluoromethanesulphonate (14.7ml) was added. The ice bath was removed, and the solution was heated at reflux under nitrogen for 45 mins. After cooling and evaporation, the residue was purified by column chromatography over 1kg of silica gel (Merck 9385) eluting with chloroform/methanol 9:1. Appropriate fractions were cooled and evaporated to afford a crude residue. This was fractionally crystallized from the minimum of hot methanol (c.1200ml) to afford the title compound (6.32g) as white crystals. ¹H NMR (d⁶DMSO) δ 11.36 (1H,bs), 7.50-8.00 (6H,m), 6.20 (1H,t), 5.46 (2H,m), 4.62 (2H, m), 3.48 (1H, m), 3.25 (1H, m).

INTERMEDIATE 3

(±)-(cis)-4-Amino-1-(2-benzovloxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-one

Method (a)

A suspension of cytosine (20.705g) and ammonium sulphate (few mgs) in hexamethyldisilazane (110ml) was stirred and heated at reflux for 2^{1/2}h, under nitrogen. Solvent was removed by evaporation, and the residual solid was dissolved in dry acetonitrile (350ml). This solution was transferred using flexible needle techniques into a stirred, ice-chilled solution of 5-methoxy-1,3-oxathiolane-2-methanol, benzoate (Intermediate I) (43.57g) in acetonitrile (650ml) under nitrogen. Trimethylsilyl trifluoromethanesulphonate (33ml) was added, the solution was allowed to warm to ambient temperature (1^{1/2}h) then heated to reflux for an overnight period. The residue mixture was concentrated, diluted with saturated aqueous sodium bicarbonate solution (500ml),

then extracted with ethyl acetate (3x500ml). The combined extracts were washed with water (2x250ml) and brine (250ml) dried (MgSO_4) then evaporated to a foam which was subjected to column chromatography on silica (600g, merck 7734), eluted with ethyl acetate-methanol mixtures to give a mixture of anomers (ca 1:1 31.59g). The mixture was crystallised from water (45ml) and ethanol (9.0ml) to give a solid (10.23g) which was recrystallised from ethanol (120ml) and water (30ml) to give the title product as a white solid (9.26g); λ_{max} (MeOH) 229.4mm ($\text{E}^{1\%}$ 610); 272.4mm ($\text{E}^{1\%}$

1cm 1cm
293); ^1H NMR (DMSO d_6) δ 3.14 (1H), 3.50 (1H), 4.07 (2H), 5.52 (1H), 5.66 (1H), 6.28 (1H), 7.22 (2H), 7.56 (2H), 7.72 (2H), 8.10 (2H).

Method (b)

Phosphorus oxychloride (7.0ml) was added dropwise to a stirred, ice-cooled suspension of 1,2,4-triazole (11.65g) in acetonitrile (120ml) then, keeping the internal temperature below 15°C, triethylamine (22.7ml) was added dropwise. After 10 min a solution of (\pm)-cis -1-(2-benzoyloxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2,4-dione (Intermediate 2) (6.27g) in acetonitrile (330ml) was slowly added. Stirring was then continued at room temperature overnight. The mixture was cooled by means of an ice bath and triethylamine (30ml) was slowly added followed by water (21ml). The resultant solution was evaporated, and the residue was partitioned between saturated sodium bicarbonate solution (400ml) and chloroform (3x200ml). The combined chloroform extracts were dried and magnesium sulphate, filtered and evaporated to give a crude residue (9.7g). The residue was dissolved in 1,4-dioxan (240ml) and concentrated aqueous ammonia solution (s.g 0.880, 50ml) was added. After 1.5h the solution was evaporated and the residue dissolved in methanol. This caused precipitation of a solid, which was filtered off. The mother liquors were purified by column chromatography over silica gel (Merck 9385, 600g). Appropriate fractions were pooled and evaporated to give the title compound as a fawn solid (2.18g), identical to that obtained by Method (a).

Example :

(2S)-(cis)-4-Amino-1-(2-hydroxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-one

A suspension of (cis)-4-amino-1-(2-benzoyloxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-one (Intermediate 3) (8.19g) and Amberlite IRA-400 (OH) resin (8.24g) in methanol (250ml) was stirred and heated to reflux for 18/h. Solids were removed by filtration then washed with methanol. The combined filtrates were evaporated. The residue was triturated with ethyl acetate (80ml). The resulting white solid was collected by filtration to give the title product (5.09g), 1H NMR (DMSO-d₆) 3.04 (1H), 3.40 (1H), 3.73 (2H), 5.18 (1H), 5.29 (1H), 5.73 (1H), 6.21 (1H), 7.19 (2H), 7.81 (1H).

Example 2: Chiral HPLC separation of the enantiomers of (2S)-(cis)-4-Amino-1-(2-hydroxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-one.

(A) The racemic product of Example 1 (25mg) was subjected to preparative HPLC under the following conditions:

Column: Merck Hibar cellulose triacetate, 250 x 10mm, 10 μ
Eluant: ethanol;
Flow: 3ml/min;
Detection: uv, 270nm;
Temperature: ambient.

Evaporation of the appropriate pooled fractions gave (2R,cis)-4-amino-1-(2-hydroxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-one (6.8mg, e.e. ca 100%) and (2S,cis)-4-amino-1-(2-hydroxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-one (3.6mg, e.e. ca 90%).

(B) The racemic product of Example 1 (26mg) was subjected to preparative HPLC under the following conditions:

Column: ASTEC cyclobond I acetyl, 250x4.6mm;
Eluant: 0.2% triethylammonium acetate (prepared by adding glacial acetic acid to 0.2% triethylamine in water, to a final pH 7.2);
Flow: 2ml/min;
Detection: uv, 300nm;
Temperature: ambient.

Evaporation of appropriate fractions gave crude (2R,cis)-4-amino-1-(2-hydroxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-one (25mg) and crude (2S,cis)-4-amino-1-(2-hydroxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-one (17mg). These fractions were separately subjected to further preparative HPLC run under the following conditions:

Column: ASTEC cyclobond I acetyl, 250x4.6mm
Eluent: 15mM ammonium acetate., pH 6.8;
Flow: 0.5ml/min
Detection: uv, 300nm;
Temperature: 5°.

Evaporation of the appropriate pooled fractions gave (2R, Cis)-4-amino-1-(2-hydroxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-one (5.0mg, e.e ca. 91%) and (2S,cis)-4-amino-1-(2-hydroxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-one (7.6mg, e.e. ca 96%).

Example 3

(-)-cis-4-Amino-1-(2-hydroxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-one

(i) (±)-cis-4-Amino-1-(2-hydroxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidinone, dihydrogenphosphate, ammonium salt

A cold (0°), stirred suspension of (±)-cis-4-amino-1-(2-hydroxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-one (1.00g) (Example 1) in dry trimethylphosphate (20ml) was treated with phosphorous oxychloride (2.44ml), and the mixture was stirred at 0° for 35 mins then quenched in ice water (60g). The cold mixture was adjusted to pH 2.5 by the addition of aqueous N-sodium hydroxide, then applied to a charcoal column (10g, DARCO), which was eluted with water, then ethanol-aqueous ammonia. Fractions containing crude monophosphate were combined and concentrated. The resulting solution was applied to a column containing 25g of DEAE Sephadex A-25 (HCO₃⁻ form). Elution was performed with a gradient of water (120ml) to 0.1 M-NH₄HCO₃ (240ml), then 0.2, 0.3 and 4M

NH_4HCO_3 (120, 240, 400ml respectively). Appropriate fractions were combined and concentrated. The residual solution was diluted with water (40ml) and freeze-dried to give the title product as a white solid (1.37g): λ_{max} (pH6 buffer), 271.0nm ($E^{1\%}_{\text{cm}} 190$); ^1H NMR (D_2O) δ 3.23 (1H), 3.55 (1H), 4.0-4.2 (2H), 5.43 (1H), 6.07 (1H), 6.33 (1H), 8.08 (1H).

(ii) (2R, cis)-4-Amino-1-(2-hydroxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-one and (2S, cis)-4-Amino-1-(2-hydroxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-one

$5'$ -Nucleotidase (from *cretalus atrox* venom) [EC 3.1.3.5] (50mg at 17 units/mg) was added to a solution of (\pm)-*cis*-4-amino-(2-hydroxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-one, $5'$ -dihydrogenphosphate, ammonium salt (1.35g) in buffer (30ml, prepared from glycine (526mg) and magnesium chloride (190mg) in water (100ml)) and the mixture was incubated at 37° for 2.5h. More enzyme (20mg) was added, and incubation was continued for a further 3.5h. The resulting mixture was applied to a column of DEAE Sephadex A-25(HCO_3 -form). Elution was performed with water (160ml), then 0.1, 0.2, 0.3 and 0.4M NH_4HCO_3 (200ml each). Appropriate fractions containing the first eluted component were combined and evaporated, the residue was subjected to column chromatography on silica (60g, Merck 7734), eluted with chloroform-methanol mixtures. Evaporation of the appropriate fractions from methanol-ethyl acetate gave (2R, *Cis*)-4-amino-1-(2-hydroxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-one as a white solid (0.30g) - $[\alpha]_D^{21} +137^\circ$ (c.1.04 MeOH); ^1H NMR (DMSO) δ 3.04 (1H), 3.40 (1H), 3.73 (2H), 5.18 (1H), 5.29 (1H), 5.73 (1H), 6.21 (1H), 7.19 (2H), 7.81 (1H).

Appropriate fractions from the Sephadex column, containing the second eluted component, were combined and evaporated. The residue was dissolved in water (30ml), treated with alkaline phosphatase (from *Escherichia coli*) [EC 3.1.3.1] (1.5ml at 416units/ml), then incubated at 37° for one hour. Solvent was removed by evaporation and the residue was subjected to column chromatography on silica (60g, Merck 7734), eluted with chloroform-methanol mixtures. Evaporation of the appropriate fractions from methanol-ethyl acetate

gave (2S,2S)-4-amino-1-(2-hydroxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-one as a white solid (0.32g); $[\alpha]_D^{25} = -132^0$ (c.1.08, MeOH); ^1H NMR (DMSO) δ 3.04 (1H), 3.40 (1H), 3.73 (2H), 5.18 (1H), 5.29 (1H), 5.73 (1H), 6.21 (1H), 7.19 (2H), 7.81 (1H).

Example 4

(-)-cis-4-Amino-1-(2-hydroxymethyl-1,3-oxathiolan-5-yl)-(1H)-pyrimidin-2-one

(i) Three 50ml flasks of nutrient broth (Oxoid Ltd) were innoculated with a loopful each of Escherichia coli (ATCC 23848) scraped from a Nutrient Agar plate. The flasks were incubated overnight at 37^0C with shaking at 250 rev/min and then each flask was used to innoculate 4l of CDD medium (glutamic acid, 3g/l; MgSO_4 , 0.3g/l; K_2SO_4 , 2.5g/l; NaCl, 2.3g/l, $\text{Na}_2\text{HPO}_4 \cdot 2\text{H}_2\text{O}$, 1.1g/l, $\text{NaH}_2\text{PO}_4 \cdot 2\text{H}_2\text{O}$ 0.6g/l cytidine, 1.2g/l) in a seven litre fermenter. The cultures were fermented at 750 rev/min, 37^0C with aeration at 4l/min. After growth for 24hrs the cells were collected by centrifugation (5000g, 30 minutes) to yield 72g wet weight. The cell pellet was resuspended in 300ml of 20mM Tris HCl buffer (pH 7.5) and disrupted by sonication (4 x 45 seconds). The cell debris was removed by centrifugation (30,000 g, 30 minutes) and the protein in the supernatant was precipitated by addition of ammonium sulphate to 75% saturation. The precipitate was collected by centrifugation (30,000g, 30 minutes) and the pellet was resuspended in 25ml of HEPES buffer (100mM, pH 7.0) containing ammonium sulphate (75% saturation). Enzyme solution was prepared by centrifugation at 12,000 rpm for 30 mins. The supernatant was discarded and the pellet dissolved in Tris HCl buffer (pH 7.0; 100mM) to the original volume.

(ii) The product of Example 1 (115mg was dissolved in water (100ml), and stirred. Enzyme solution (0.5ml) was added, and the mixture was maintained at a constant pH by the continual addition of HCl (25mM). The conversion was monitored by chiral HPLC, which showed that the (-) enantiomer of the substrate was preferentially deaminated. After 22hr the (+) enantiomer of the substrate (RT 12.5min) had been

completely removed, and the solution was adjusted to pH 10.5 by the addition of conc. sodium hydroxide.

The solution produced above was eluted through a column of QAE Sephadex (A25; Pharmacia; 30X 1.6cm), pre-equilibrated to pH11. The column was washed with water (200ml) and then with HCl (0.1M). Fractions (40ml) were taken, and analysed by reversed phase HPLC. Fractions 5-13, containing the unreacted (-) enantiomer of the substrate, were combined and adjusted to pH 7.5 with HCl.

Fraction 47, containing deaminated product, was adjusted to pH7.5 with dil. NaOH. Analysis by chiral HPLC showed that this material was a mixture, consisting of one enantiomer (RT 10.2min) as the major component with the other enantiomer (RT 8.5min) as a minor component (e.e ca 90%).

(iii) Stage (ii) above was repeated on a larger scale. The compound of Example 1 (363mg) in 250ml of water was incubated with enzyme solution (0.5ml), prepared as in Stage (i). Further aliquots (0.5ml) of enzyme were added after 18 and 47 hrs. The reaction mixture was stirred for 70hr., then left standing for a further 64hr. Analysis by chiral hplc indicated that the (+) enantiomer of the substrate had been completely deaminated, and the resulting solution was adjusted to pH10.5 with NaOH.

The solution above was loaded onto the same QAE column, and eluted as in stage (i). Fractions 2-6, containing a mixture of the residual substrate and deaminated product, were bulked. Fractions 7-13, containing the residual substrate ((-) enantiomer), were bulked and adjusted to pH7.5. Fractions 25-26, containing deaminated product, were bulked and neutralised.

Fractions 2-6 above were re-eluted through the same QAE column. Fractions 3-11 from this second column contained unreacted substrate ((-) enantiomer). Fraction 70 contained the deaminated product.

(iv) The resolved substrate fractions from stage (ii) and (iii) were combined and adjusted to pH7.5. This solution was eluted through a column of XAD-16 (40x2.4cm), packed in water. The column was washed with water, and then eluted with acetone: water (1:4 v/v).

Fractions containing the (-) enantiomer of BCH 189 were bulked and freeze-dried to give a white powder (190mg).

The HPLC methods used above were as follows:-

1. Reversed Phase analytical HPLC

Column	:	Capital Cartridge
		Spherisorb CDS-2 (5μm)
		150x4.6mm
Eluant	:	Ammonium dihydrogen phosphate (50mM) + 5% MeCN
Flow	:	1.5ml/min
Detection	:	UV, 270nm
Retention Times	:	BCH 189 5.5min
	:	deaminated BCH -189 8.1min

2. Chiral analytical HPLC

Column	:	Cyclobond I Acetyl
		250x4.6mm
Eluant	:	0.2% Triethylammonium acetate (pH7.2)
Flow	:	1.0ml/min
Detection	:	UV, 270nm
Retention Times	:	BCH 189 11.0 and 12.5min
	:	deaminated BCH-189 8.5 and 10.2 min

(The bioconversion was followed by monitoring the loss of the peak at 12.5min., and accumulated of product at 10.2min).

Example 5

(-)-cis-4-Amino-1-(2-hydroxymethyl-1,3-oxathiolan-5-yl)-(1H)-
pyrimidin-2-one

A loopful of *E. coli* B cells (ATCC 32848) scraped from a well grown nutrient agar plate was used to inoculate two Florence flasks each containing 250ml of nutrient broth. The culture was incubated at 37° with shaking (250 rev/min, 5cm throw) for 18 hours. This was then used to inoculate 40L of CDD-medium, with cytidine, in a 70L fermenter.

Conditions for the fermentation were as follows : 40L/min aeration, 750 rev/min stirring speed at a temperature of 37°C. Three Rushton impellers were fitted to the fermenter. The fermentation was run for 18 hours before harvest using a Sharples continuous centrifuge. The cell paste (150g net weight) was frozen at -20°C prior to cell breakage.

ODD Medium

	g/L
L-Glutamic acid	3
MgSO ₄	0.2
K ₂ SC ₄	2.5
NaCl	2.3
Na ₂ HPO ₄	1.1
NaH ₂ PO ₄	0.6

Made with distilled water. Sterilised at 121°C for 30 minutes. Cytidine (1.2g/L) was filter sterilised and added prior to inoculation.

The frozen cell paste (150g) was thawed and suspended in 750ml of 100mM Hepes (N-[2-hydroxyethyl]piperazine-N'-(2-ethane sulphonic acid)) buffer (pH 7.5) containing 1mM ethylenediaminetetraacetic acid (sodium salt) and 1mM dithiothreitol (disruption buffer). The cells were disrupted by passing the suspension through a Manton-Gaulin homogeniser operating at 7500psi. This was carried out three times with cooling of the suspension to approximately 5°C after each pass through the homogeniser. The homogenate was clarified by centrifugation (1400g; 60min). The cytidine deaminase activity was adsorbed onto a Q-Sepharose column (490mg bed volume) pre-equilibrated with 50mM Tris (hydroxymethyl) methylamine (pH 7.5) containing 1mM sodium chloride. The pooled active fractions (210ml) were added onto a phenyl-Sepharose column (490ml bed volume), pre-equilibrated with the sample buffer containing 3.2M ammonium sulphate. The bound enzyme was eluted by a gradient of decreasing ammonium sulphate concentration. Fractions containing the cytidine

deaminase activity were pooled (695ml) and the partially purified enzyme was then precipitated with 80% ammonium sulphate. After centrifugation (1400g; 60min) the pellet was resuspended in 54ml of supernatant from the above and stored at 4°C.

6.2ml of this solution was centrifuged (18000g, 90min) and the pellet was dissolved in 24ml of 0.5M potassium phosphate buffer (pH 7.5). The suspension was dialysed overnight against 1M potassium phosphate buffer (pH 7.5). The retentate (20ml) was then diluted with an equal volume of distilled water. To 35ml of this solution dry Supergit C beads were added (1g) and the mixture was allowed to stand at room temperature 150-300 hours (determined by measuring residual cytidine deaminase activity in solution). The immobilised enzyme was washed with 100mM Tris/HCl buffer (pH 7.0) containing 1mM EDTA, 1mM DTT, 0.5M NaCl and 500ppm p-hydroxybenzoic acid ethyl ester (storage buffer). The immobilised enzyme (2.7g wet weight) was stored in this buffer at 4°C until required for biotransformation.

The product of Example 1 (3g) was dissolved in 500ml of distilled water in a magnetically stirred 1L flask. The bioconversion was carried out at 32°C in a pH-stat. The pH was maintained constant at 7.0 by addition of 1M acetic acid. 1g Wet weight of the immobilised enzyme beads was washed with distilled water prior to start of the reaction. The conversion was monitored by chiral-HPLC which showed that the (+) enantiomer of the substrate was preferentially deaminated. At the end of the reaction (72h) the enzyme beads were filtered off and the filtrate was used for isolation of the desired (-)-enantiomer.

The pH of the reaction mixture was adjusted to pH 10.5 using ammonia solution (1M) and the solution applied to Duolite A113 super resin in the OH cycle (50ml; 0.4 bed volumes per hour). The uridine analogue adsorbed to the resin and the (-)-enantiomer passed straight through. Any (-)-enantiomer still on the resin was removed by washing with 0.04% ammonia solution (2 bed volumes; flow rate 0.8 bed volumes/hour).

The pH of the spent solutions and washes (600ml) was adjusted to pH 7.0 with concentrated sulphuric acid and the solution applied to XAD16 resin (50ml; flow rate 1.4 bed volumes per hour). The

column was washed with distilled water (2.5 bed volumes, flow rate 2 bed volumes per hour) and the (-)-enantiomer eluted with acetone:water 1:3 (flow rate 1.5 bed volumes per hour).

The bulk (-)-enantiomer containing fraction (4 bed volumes) was concentrated on a Buchni evaporator to a small volume before filtration through a No. 3 glass sinter. The filtered solution was freeze dried to yield 1.2g of title product identical to that obtained in Example 4.

Example 6

Tablet Formulations

A. The following formulation is prepared by wet granulation of the ingredients with a solution of providone in water, drying and screening, followed by addition of magnesium stearate and compression.

	mg/tablet
(a) Active ingredient	250
(b) Lactose B.P.	210
(c) Providone B.P.	15
(d) Sodium Starch Glycolate	20
(e) Magnesium Stearate	5
	—
	500

B. The following formulation is prepared by direct compression; the lactose is of the direct compression type.

	mg/tablet
Active Ingredient	250
Lactose	145
Avicel	100
Magnesium Stearate	5
	—
	500

C. (Controlled Release Formulation). The formulation is prepared by wet granulation of the ingredients (below) with a solution of providone in water, drying and screening followed by the addition of magnesium stearate and compression.

	mg/tablet
(a) Active ingredient	500
(b) Hydroxypropylmethylcellulose (Methocel K4M Premium)	112
(c) Lactose B.P.	53
(d) Providone B.P.	28
(e) Magnesium Stearate	7
	—
	700

Example 7

Capsule Formulation

A capsule formulation is prepared by admixing the ingredients below and filling into a two-part hard gelatin capsule.

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	mg/capsule
Active ingredient	125
Lactose	72.5
Avicel	50
Magnesium Stearate	2.5
	—
	250

Example 3

Injectable Formulation

Active ingredient 0.200g

Sodium hydroxide solution, 0.1M q.s. to a pH of about 11.

Sterile water q.s. to 10ml

The active ingredient is suspended in some of the water (which may be warmed) and the pH adjusted to about 11 with a solution of sodium hydroxide. The batch is then made up to volume and filtered through a sterilizing grade membrane filter into a sterile 10ml glass vial and sealed with sterile closures and overseals.

Example 3

	mg/suppository
Active ingredient	250
Hard Fat, B.P.	1770
	—
	2020

One-fifth of the hard fat is melted in a steam-jacketed pan at 45°C maximum. The active ingredient is sifted through a 200μm sieve and added to the molten base with mixing, using a high shear

stirrer, until a smooth dispersion is achieved. Maintaining the mixture at 45°C, the remaining hard fat is added to the suspension and stirred to ensure a homogenous mix. The entire suspension is passed through a 250µm stainless steel screen and, with continuous stirring, is allowed to cool to 40°C. At a temperature of 38°C to 40°C, 2.02g of the mixture is filled into suitable, 0ml plastic moulds. The suppositories are allowed to cool to room temperature.

Example 10

Biological Activity

Antiviral Activity

Antiviral activity of the compounds of Example 2 were determined against three strains of HIV-1 and one strain of HIV-2 in the following cell lines.

JM cells, a semi-mature T-cell line derived from a patient with lymphoblastic leukaemia, infected with HIV-1 strain GB8.

C8166 cells, a human T-lymphoblastoid cell line, infected with HIV-1 strain RF.

MT-4 cells, a human T-cell leukaemia cell line, infected with HIV-1 strain RF.

CEM cells, a human T-lymphoblastoid cell line, infected with HIV-1 strains RF and U455, or HIV-2 strain ROD.

Antiviral activities in C8166, JM, and CEM cells were determined by inhibition of syncytium formation (Tochikura et al *Virology*, 164, 542-546) and in MT-4 cells by inhibition of formazan conversion (Baba et al; (1987) *Biochem Biophys Res Commun.*, 142, 128-134; Mossman (1983) *J. Immun Meth.* 65, 55-57). Antiviral activities were also determined by analysing the amount of HIV p24 antigen synthesised in the presence and absence of enantiomers.

The results are shown in Tables 1 and 2 below:-

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Table 1

		50% Antiviral Activity (μ g/ml)					
Assay	Formazan	inhibition of syncytium formation					
Cells	MT-4	CEM	CEM	CEM	JM	C8166	
Virus (HIV-1)	HIV-1 RF	HIV-2 ROD	HIV-1 RF	HIV-1 U455	HIV-1GB8	HIV-1 RF	
(+)-enantiomer	0.28	0.045	0.07	0.006	0.03	0.05	
(-)-enantiomer	0.20	0.055	0.04	0.008	0.05	0.01	

Table 2

Inhibition of HIV p24 Synthesis

50% Inhibition HIV p24 Synthesis (μ g/ml)			
Cells	C8166	JM	MT-4
Virus	RF	GB8	RF
(+)-enantiomer	0.021	0.033	0.0008
(-)-enantiomer	0.016	0.016	0.0004

3. Cytotoxicity

The cytotoxicities of the compounds of Example 2, the racemic compound (BCH-189; Example 1) and a 50/50 mixture of the two enantiomers were determined in five CD4 cell lines: H9, JM, CEM, C8166 and U937.

Compounds for test were serially diluted from 100 μ g/ml to 0.3 μ g/ml (final concentrations) in 96 well microtitre plates. 3.6x10³ cells were inoculated into each well of the plates including drug-free controls. After incubation at 37°C for 5 days, the viable cell count was determined by removing a sample of cell suspension and counting trypan blue excluding cells in a haemocytometer.

The results are shown in Table 3.

Table 3

Cytotoxicity

Compound	50% Cytotoxicity (μ g/ml)				
	CEM cells	JM cells	H9 cells	U937 cells	C8166 cells
(+)-enantiomer	1	1.5	2	4	35
(-)-enantiomer	>100	30	>100	>100	>100
BCH-189	3	3.5	8	15	90
1:1 Mix of	2.5	ND*	ND	ND	ND

ND = Not Determined

Claims

1. (-)-cis-4-Amino-1-(2-hydroxymethyl-1,3-oxathiolane-5-yl)-(1*H*)-pyrimidin-2-one or a pharmaceutically acceptable derivative thereof.
2. A compound as claimed in Claim 1 substantially free of the corresponding (+)-enantiomer.
3. A compound as claimed in Claim 1 or Claim 2 wherein the (+)-enantiomer is present in an amount of no more than about 5% w/w.
4. A compound as claimed in any one of Claims 1 to 3 wherein the (-)-enantiomer is present in an amount of no more than about 2% w/w.
5. A compound as claimed in any one of Claims 1 to 4 wherein the (-)-enantiomer is present in an amount of less than about 1% w/w.
6. A compound as claimed in Claim 1 in substantially pure form.
7. A pharmaceutical composition comprising a compound as claimed in any one of Claims 1 to 6 together with a pharmaceutically acceptable carrier therefor.
8. A compound as claimed in any one of Claims 1 to 6 for use in therapy.
9. Use of a compound as claimed in any one of Claims 1 to 6 for the manufacture of a medicament for the treatment of a viral infection.
10. A method for the treatment of a mammal, including man, suffering from or susceptible to viral infection comprising administration of an effective amount of a compound as claimed in any one of Claims 1 to 6.

11. A method for the preparation of a compound as claimed in any one of Claims 1 to 6 which comprises separation of the (-)-enantiomer from a mixture also containing the (+)-enantiomer.
12. A method as claimed in Claim 11 wherein the mixture of compounds is a racemic mixture.
13. A method as claimed in Claim 11 or Claim 12 the separation is effected by chiral HPLC.
14. A method as claimed in Claim 13 wherein the HPLC employs as a stationary phase acetylated β -cyclodextrin or cellulose triacetate.
15. A method as claimed in Claim 11 or Claim 12 wherein the separation is effected by enzyme-mediated enantioselective catabolism.
16. A method is claimed in Claim 15 wherein the enzyme is employed in immobilised form.
17. A method as claimed in Claim 15 or Claim 16 wherein the enzyme is cytidine deaminase.
18. A method as claimed in Claim 15 or Claim 16 wherein the enzyme is a 5'-nucleotidase.

INTERNATIONAL SEARCH REPORT

International Application No. PCT/GB 91/00706

I. CLASSIFICATION & SUBJECT MATTER (if several classification symbols apply, indicate all) *

According to International Patent Classification (IPC) or to both National Classification and IPC

IPC 5 C 07 D 411/04, A 61 K 31/505

II. FIELDS SEARCHED

Minimum Documentation Searched *

Classification System | Classification Symbols

IPC⁵

C 07 D 411/04, C 07 D 405/04

Documentation Searched other than Minimum Documentation
to the Extent that such Documents are Included in the Fields Searched *

III. DOCUMENTS CONSIDERED TO BE RELEVANT *

Category * Citation of Document, ** with indication, where appropriate, of the relevant passages *** Relevant to Claim No. ***

A EP, A, 0337713 (I.A.F.)
18 October 1989
see pages 1-5,11; claims

1,6-9

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P,X EP, A, 0382526 (I.A.F.)
16 August 1990
see page 17, example 7; page 27;
claims

1,6-9

- * Special categories of cited documents: 10
- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the International filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the International filing date but later than the priority date claimed

- "T" later document published after the International filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step
- "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
- "E" document member of the same patent family

IV. CERTIFICATION

Date of the Actual Completion of the International Search

31st July 1991

Date of Mailing of this International Search Report

- 2. 09. 91

International Searching Authority

EUROPEAN PATENT OFFICE

Signature of Authorized Officer

M. PEIS

M. Peis

VI. OBSERVATIONS WHERE CERTAIN CLAIMS WERE FOUND UNSEARCHABLE:

This International search report has not been established in respect of certain claims under Article 17(2) (a) for the following reasons:
1. Claim numbers 10, because they relate to subject matter not required to be searched by this Authority, namely:

Pls. see Rule 39.1(iv) - PCT:

Methods for treatment of the human or animal body by therapy or surgery as well as diagnostic methods.

2. Claim numbers , because they relate to parts of the International application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. Claim numbers , because they are dependent claims and are not drafted in accordance with the second and third sentences of PCT Rule 6.4(a).

VI. OBSERVATIONS WHERE UNITY OF INVENTION IS LACKING:

This International Searching Authority found multiple inventions in this International application as follows:

1. As all required additional search fees were timely paid by the applicant, this International search report covers all searchable claims of the International application.

2. As only some of the required additional search fees were timely paid by the applicant, this International search report covers only these claims of the International application for which fees were paid, specifically claims:

3. No required additional search fees were timely paid by the applicant. Consequently, this International search report is restricted to the invention first mentioned in the claims; it is covered by claim numbers:

4. As all searchable claims could be searched without effort justifying an additional fee, the International Searching Authority did not make payment of any additional fee.

Remarks on Patent:

- The additional search fees were accompanied by applicant's patent.
- No patent accompanied the payment of additional search fees.

ANNEX TO THE INTERNATIONAL SEARCH REPORT
ON INTERNATIONAL PATENT APPLICATION NO.

GB 9100706
SA 47177

This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report. The members are as contained in the European Patent Office EDP file on 27/08/91. The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

Patent document cited in search report	Publication date	Patent family members)		Publication date
EP-A- 0337713	18-10-89	AU-A-	3264489	12-10-89
		JP-A-	1316375	21-12-89
EP-A- 0382526	16-08-90	AU-A-	4920190	16-08-90
		CA-A-	2009637	08-08-90
		JP-A-	3007282	14-01-91